

Utilizing the OHAUS Guardian 5000 Hotplate Stirrer to incubate a sample during Bradford Protein Assay

For labs working with proteins, it is imperative that they are able to determine protein concentration within a given solution. One way to determine this is through the use of a Bradford assay. The assay involves Coomassie Brilliant Blue G-250 dye, which causes a color change when interacting with the sample under certain conditions. This color change can then be correlated with a numerical value through an absorbance reader. Bradford Assays are a common practice as they are both rapid and highly sensitive.

### Preparation of Protein Solution

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1. Create a cell suspension by homogenizing the tissue sample using the [OHAUS HT Lysing Homogenizer](#).
2. Remove a 10ml sample from the well-mixed cell suspension, and centrifuge it at 8,000g for 15 minutes with the [OHAUS Frontier 5000 Series Multi Pro](#). Discard supernatant.
3. Wash pellets with 0.9% m/v NaCl, spin again, and resuspend in 2ml 0.2 M NaOH in a 28mm vial.
4. Install the base plate and the 28mm vial block on the [OHAUS Guardian 5000 Hotplate Stirrer](#).
5. Turn on the hotplate and set the device to 100 °C. The hot top light will illuminate when the device gets above 40 °C.
6. Incubate for 10 minutes at 100 °C and remove the vial from the hotplate.
7. After cooling, centrifuge at 8,000g for 15 minutes. Collect supernatant for Bradford assay.

### Bradford Protein Assay

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1. In 96 well plate, carefully pipette 100µL protein assay dye 1x in each well. Take caution to not expose the assay dye to a direct light source for prolonged periods of time.
2. Add 20µL sample, mix well by pipette, then seal the plate.
3. Incubate at room temperature ~5 min. There should be a noticeable color change of the solution as the reaction takes place.
4. Read the absorbance with a plate reader at 595nm.

## OHAUS Products Used Within This Procedure

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OHAUS HT Lysing  
Homogenizer



OHAUS Frontier 5000 Series  
Multi Pro



OHAUS Guardian 5000  
Hotplate Stirrer